

Hantavirus Awareness & Prevention Guide

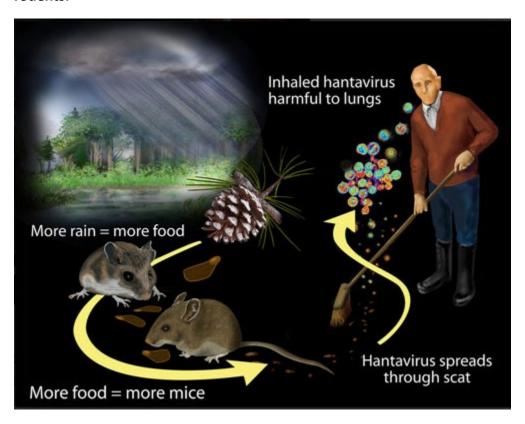
For UC Riverside Personnel Working with Field Rodents

What is Hantavirus?

Hantavirus is a serious and potentially fatal illness transmitted by infected rodents. Humans can become infected through:

- Inhaling dust contaminated with rodent urine, droppings, or saliva
- Direct contact with rodent bodily fluids
- (Rarely) through rodent bites

While dogs and cats are not direct carriers, they may inadvertently expose humans to infected rodents.



Primary Carrier in California: Deer Mouse (Peromyscus maniculatus)

- Size: Comparable to a house mouse
- Color: Pale gray to reddish-brown with a white belly and feet
- Tail: Bi-colored and slightly shorter than the body
- Habitat: Found in forests, grasslands, chaparral, and brushy areas throughout California

Who's at Risk at UCR?

- Field researchers or students handling or trapping wild rodents
- Staff cleaning field stations, barns, attics, or storage areas
- Maintenance personnel working in crawl spaces or unused buildings
- Campers or students working in remote or rural field locations

Recognizing Hantavirus Symptoms

Symptoms typically appear 1–6 weeks after exposure and may include:

- High fever (101–104°F)
- Muscle aches, chills, and headache
- Abdominal, joint, or back pain
- Nausea and vomiting
- Difficulty breathing due to fluid in the lungs

Hantavirus Pulmonary Syndrome (HPS) can be fatal. If symptoms develop after potential exposure, seek immediate medical care and inform your provider of the rodent exposure.

Prevention Measures

Rodent Elimination

- Use snap traps baited with peanut butter for at least one week
- Treat affected areas for fleas as needed

Use tamper-resistant bait stations in accordance with UC policy

Disposal Protocol

- Spray rodents and contaminated materials with disinfectant before handling
- Use gloves or an inverted plastic bag to handle carcasses
- Seal waste in a plastic bag and dispose of in a rodent-proof container

Safe Cleaning Practices

Before entering a rodent-infested area:

- Air out the area for at least 30 minutes
- Wear gloves, eye protection, N95 or P100 mask, and protective clothing

For cleaning:

- Use a bleach solution (1 part bleach to 9 parts water) or an EPA-registered disinfectant
- Spray all contaminated materials before and after handling—do not sweep or vacuum
- Mop floors, disinfect hard surfaces, and wash bedding/clothing in hot water

UCR Fieldwork Safety Guidelines

- Attend EH&S Field Safety & Hantavirus Awareness Training
- Wear HEPA-filtered respirators in enclosed or high-risk rodent areas (contact EH&S medical clearance and for fitting)
- Decontaminate reusable PPE and equipment after field use
- Do not eat, drink, or touch your face when handling rodents or cleaning traps

Working with Laboratory and Wild Rodents

- Lab colonies of deer mice must test negative for hantavirus and be re-tested regularly (check with your facility veterinarian)
- Do not mix wild-caught mice with lab-reared colonies
- Isolate wild-caught rodents until cleared by testing

 Field biologists must treat wild deer mice as potentially infected and wear EH&Sapproved HEPA respirators when handling or cleaning

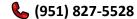
When cleaning rodent-infested structures:

- Wear a fitted HEPA respirator and gloves
- Ventilate the structure for at least 24 hours
- Thoroughly spray all surfaces with disinfectant
- Disinfect gloves before removal and wash hands afterward
- Avoid sleeping in contaminated areas

Need Help or More Information?

Contact UCR Environmental Health & Safety (EH&S):

ehs@ucr.edu



ehs.ucr.edu

https://www.cdph.ca.gov/Programs/CID/DCDC/pages/hantaviruspulmonarysyndrome.aspx#: ~:text=How%20can%20l%20help%20p,containers%20to%20keep%20mice%20away.